

Direct Experimental Characterization of the Ferrier Glycosyl Cation in the Gas Phase

Kim Greis, Carla Kirschbaum, Sabrina Lechnitz, Sandy Gewinner, Wieland Schöllkopf, Gert von Helden, Gerard Meijer, Peter H. Seeberger, and Kevin Pagel*

Cite This: *Org. Lett.* 2020, 22, 8916–8919

Read Online

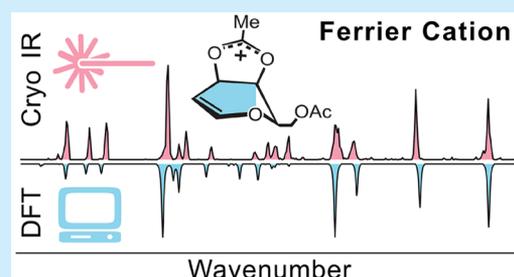
ACCESS |

Metrics & More

Article Recommendations

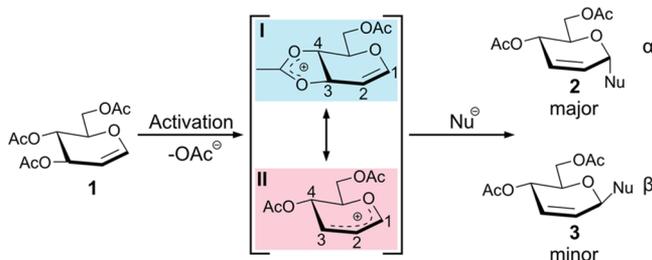
Supporting Information

ABSTRACT: The Ferrier rearrangement reaction is crucial for the synthesis of pharmaceuticals. Although its mechanism was described more than 50 years ago, the structure of the intermediate remains elusive. Two structures have been proposed for this Ferrier glycosyl cation: a 1,2-unsaturated cation that is resonance-stabilized within the pyranose ring or a cation that is stabilized by the anchimeric assistance of a neighboring acetyl group. Using a combination of gas-phase cryogenic infrared spectroscopy in helium nanodroplets and first-principles density functional theory, we provide the first direct structural characterization of Ferrier cations. The data show that both acetylated glucal and galactal lead to glycosyl cations of the dioxolenium type.



In 1917, Emil Fischer reported a class of 1,2-unsaturated monosaccharide building blocks, so-called glycols.¹ Almost 50 years later, the rich chemistry of glycols was exploited by Robert J. Ferrier, who first accurately described the mechanism of the allylic rearrangement in protected glycols upon nucleophilic substitution, today known as the Ferrier rearrangement (Scheme 1).^{2,3} In this reaction, the C3-acetyl

Scheme 1. Ferrier Rearrangement Reaction, in Which Activation of Acetylated Glucal Leads to Ferrier Glycosyl Cations That Can Be Attacked by Nucleophiles



group in fully acetylated glycols is cleaved to form the “Ferrier” glycosyl cation that in turn is attacked by nucleophiles at the C1 position. After an allylic shift, the reaction yields 2,3-unsaturated glycosides, a class of glycosides that is relevant for the synthesis of natural products⁴ and pharmaceuticals such as 2-deoxyglycosides,⁵ C-glycopyranosyl arenes, and heteroarenes.⁶ The reaction can also be observed for glycols bearing other participating protecting groups, i.e., benzoyl.³

Due to their short-lived nature, the exact structure of the intermediate of the Ferrier rearrangement remains elusive. It has been postulated that cleavage of the C3-acetyl group is

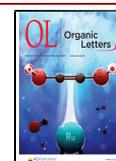
eased by neighboring group participation (anchimeric assistance) of the C4-acetyl group, leading to a dioxolenium-type ion I.⁷ However, so far no direct experimental evidence has been available to determine whether the underlying structural motif is indeed a dioxolenium ion I or a resonance-stabilized oxocarbenium ion II. Recently, a Ferrier cation with protonated acetyl groups was characterized by NMR spectroscopy in superacids. As protonation of acetyl groups drastically reduces nucleophilicity, participation cannot be observed under these conditions.⁸ In a previous study of glycosyl cations, it was shown that C4-acetyl groups, contrary to C2-acetyl groups, do not participate in superacids.⁹

In the past several years, it has been demonstrated that glycosyl cations can be isolated in the gas phase and characterized using infrared (IR) spectroscopy. In combination with density functional theory (DFT), the underlying structural motif can be unambiguously identified. Thereby, it has been shown that neighboring and remote acetyl groups can stabilize a glycosyl cation by forming a covalent bond to the anomeric carbon.^{10–14} Here, we use gas-phase cryogenic IR spectroscopy and DFT to elucidate the structure of the Ferrier glycosyl cations generated from acetylated glucal and galactal.

A detailed description of the experimental setup can be found in the [Supporting Information](#). Briefly, Ferrier cations are generated by nano-electrospray ionization (nESI) followed

Received: October 1, 2020

Published: November 5, 2020



by in-source fragmentation of sodiated glycal precursor ions. Representative mass spectra (Figure 1) reveal that fragmenta-

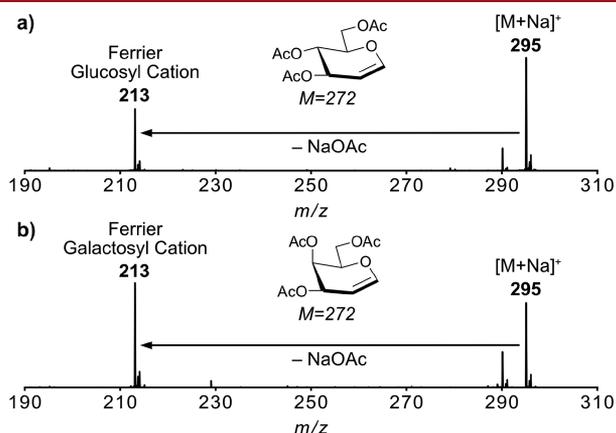


Figure 1. ESI-(+)-mass spectra of (a) 3,4,6-tri-*O*-acetyl-D-glucal and (b) 3,4,6-tri-*O*-acetyl-D-galactal, detected as $[M + Na]^+$ at m/z 295. The $[M + Na]^+$ ions undergo loss of NaOAc under in-source CID conditions leading to Ferrier cations at m/z 213. $[M + NH_4]^+$ adducts can be observed at m/z 290.

tion readily occurs for acetylated glycals, while it cannot be observed for unprotected glycals (Figures S1 and S2). This is mainly attributed to acetate being a better leaving group than hydroxide and the anchimeric assistance of neighboring acetyl groups.

The generated Ferrier cations are isolated using a quadrupole mass filter and subsequently collected in a hexapole ion trap, where the ions are cooled to 90 K. A beam of superfluid helium nanodroplets is guided through the ion trap, picking up analyte ions, cooling them to 0.4 K, and leading them to a detection region. Here, the beam of ion-doped helium droplets overlaps with an IR beam (1000–1800 cm^{-1}) that is produced by the Fritz Haber Institute free-electron laser (FHI FEL).¹⁵ The absorption of multiple resonant IR photons leads to release of the analyte ions from the droplets that are subsequently detected by a time-of-flight mass spectrometer. Plotting the ion count as a function of the IR wavenumber leads to a highly resolved IR spectrum of the analyzed ion.

To derive structural information, the experimental IR spectra were linked to quantum chemically modeled structures. The conformational space of the Ferrier cations was explored using the genetic algorithm FAFOOM that allows for sampling of rotatable bonds and ring puckers.¹⁶ Each generated structure was preoptimized by the dispersion-corrected PBE+vdW^TS density functional as implemented in FHI-aims using *light* basis set settings.^{17,18} The conformational search mainly yielded dioxolenium ions **I** and oxocarbenium ions **II**, but also dioxolenium-type structures in which the C4-acetyl group interacts remotely with the anomeric carbon (remote participation). A subset of the lowest-energy structures of each type of ions was reoptimized, and frequencies were computed at the PBE0+D3/6-311+G(d,p) level of theory using Gaussian 16.^{19–21} The computed IR spectra were normalized and scaled by a factor of 0.965. Cremer–Pople coordinates were used to assign ring puckers.^{22,23} Energetics, ring puckers, and coordinates of the reoptimized structures can be found in the Supporting Information.

The IR spectrum of the Ferrier glucosyl cation shows a complex signature that can be interpreted using the computed spectra of low-energy conformers (Figure 2a). The experimental IR signature matches the theoretical spectrum of dioxolenium-type glycosyl cation **I**, exhibiting a ⁵*E* ring pucker in which the carbonyl oxygen of the C4-acetyl group forms a covalent bond (1.53 Å) with the C3 atom (Figure 2b). The free energy of this structural motif is generally more favorable than that of oxocarbenium ions **II** that adopt a distorted ⁵*E* ring pucker without direct participation of protecting groups (Figure 2c). Generally, the most diagnostic absorption bands are found above 1500 cm^{-1} . Bands at 1506 and 1550 cm^{-1} correspond to the symmetric and antisymmetric O–C–O stretches, while the signal at 1646 cm^{-1} represents a C=C stretching vibration. The pronounced absorption band at 1750 cm^{-1} can be attributed to the C=O stretch. Features below 1300 cm^{-1} result from C–C and C–O stretching vibrations, while the bands around 1400 cm^{-1} are due to C–H bending modes. Although the absorption bands in the region below 1500 cm^{-1} are usually less diagnostic, theory fits the experiment surprisingly well. The spectral signature of the energetically unfavored (+29 $kJ mol^{-1}$) oxocarbenium-type species **II** on the other hand does not match the experimental

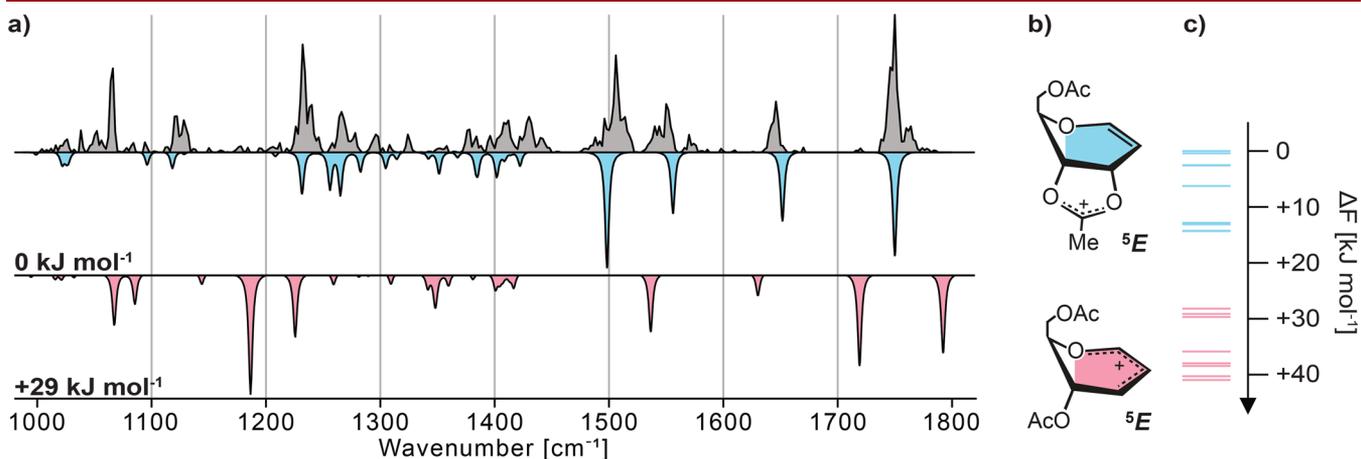


Figure 2. (a) Infrared spectra of Ferrier glucosyl cations generated from a fully acetylated glucal precursor. The experimental IR spectrum is shown as a gray trace, while computed spectra of low-energy dioxolenium (blue) and oxocarbenium (red) structures are represented in the inverted traces below. (b) Simplified representation of the low-energy structures and puckers. (c) Free energies of reoptimized low-energy structures at 90 K.

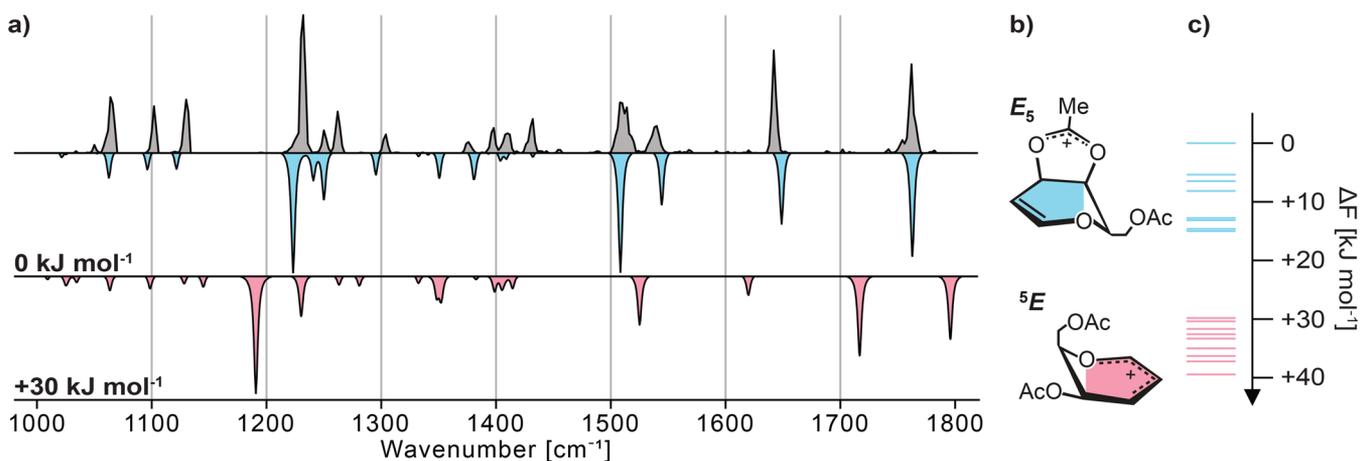


Figure 3. (a) Infrared spectra of Ferrier galactosyl cations generated from a fully acetylated galactal precursor. The experimental IR spectrum is shown as a gray trace, while computed spectra of low-energy dioxolenium (blue) and oxocarbenium (red) structures are represented in the inverted traces below. (b) Simplified representation of the low-energy structures and puckers. (c) Free energies of reoptimized low-energy structures at 90 K.

spectrum. While some signals seem to overlap, others are not present in the experimental spectrum and vice versa. In addition, none of the oxocarbenium-type species **II** matches the experimental spectrum particularly well. The spectra of the energetically less favored dioxolenium ion featuring remote participation (+49 kJ mol⁻¹) do also not correspond to the experimental spectrum (Figure S7). We can therefore conclude that the underlying structural motif of the Ferrier glucosyl cation exhibits anchimeric assistance of the C4-acetyl group to form dioxolenium-type **I** structures.

Similar results were obtained for the Ferrier galactosyl cation (Figure 3a). The computed spectrum of the dioxolenium-type **I** structure overwhelmingly matches the experimental spectrum. Besides the highly diagnostic vibrations above 1500 cm⁻¹, here also the fingerprint region from 1000 to 1300 cm⁻¹ agrees exceptionally well. Again, the dioxolenium-type **I** ion features a covalent bond (1.52 Å) between the carbonyl oxygen of the C4-acetyl group and the C3 atom and adopts an *E*₅ ring pucker (Figure 3b). The lowest-energy oxocarbenium-type **II** structure adopts a ⁵*E* ring pucker but exhibits a free energy that is 30 kJ mol⁻¹ larger than that of the lowest-energy dioxolenium-type **I** structure (Figure 3c). Not surprisingly, the spectral signatures of the oxocarbenium-type **II** structures generally match the experiment rather poorly. The Ferrier galactosyl cation adopts a dioxolenium-type **I** structure, formed by neighboring group participation of the C4-acetyl group.

In summary, we unraveled the detailed molecular structure of the key intermediate of the Ferrier rearrangement reaction. Direct experimental characterization by gas-phase cryogenic IR spectroscopy combined with first-principles density functional theory unambiguously revealed that dioxolenium-type ions are the underlying structural motif of Ferrier glycosyl cations. In cations generated from both glucal and galactal, stabilization of the positive charge at C3 via participation of the C4-acetyl group is clearly favored over delocalization of the charge within the pyranose ring. The findings provide unprecedented insights into the mechanism of the Ferrier rearrangement and showcase how such experiments can support the development of novel building blocks for the synthesis of 2,3-unsaturated glycosides.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.orglett.0c03301>.

A detailed description of the experiment, mass spectra, computed energetics, spectra, and structures (PDF)

■ AUTHOR INFORMATION

Corresponding Author

Kevin Pagel – Freie Universität Berlin, Institute of Chemistry and Biochemistry, 14195 Berlin, Germany; Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany; orcid.org/0000-0001-8054-4718; Email: kevin.pagel@fu-berlin.de

Authors

Kim Greis – Freie Universität Berlin, Institute of Chemistry and Biochemistry, 14195 Berlin, Germany; Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany

Carla Kirschbaum – Freie Universität Berlin, Institute of Chemistry and Biochemistry, 14195 Berlin, Germany; Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany

Sabrina Lechnitz – Freie Universität Berlin, Institute of Chemistry and Biochemistry, 14195 Berlin, Germany; Max Planck Institute of Colloids and Interfaces, 14476 Potsdam, Germany

Sandy Gewinner – Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany

Wieland Schöllkopf – Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany; orcid.org/0000-0003-0564-203X

Gert von Helden – Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany; orcid.org/0000-0001-7611-8740

Gerard Meijer – Fritz Haber Institute of the Max Planck Society, 14195 Berlin, Germany; orcid.org/0000-0001-9669-8340

Peter H. Seeberger – Freie Universität Berlin, Institute of Chemistry and Biochemistry, 14195 Berlin, Germany; Max

Planck Institute of Colloids and Interfaces, 14476 Potsdam, Germany

Complete contact information is available at:
<https://pubs.acs.org/10.1021/acs.orglett.0c03301>

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors acknowledge generous funding by the European Research Council, ERC-2019-CoG-863934-GlycoSpec. K.G. is grateful to the Fonds National de la Recherche (FNR), Luxembourg, for funding the project GlycoCat (13549747). C.K. is grateful for financial support by Fonds der Chemischen Industrie.

REFERENCES

- (1) Fischer, E. Über neue Reduktionsprodukte des Traubenzuckers: Glucal und Hydro-glucal. *Ber. Dtsch. Chem. Ges.* **1914**, *47*, 196.
- (2) Ferrier, R. J.; Overend, W. G.; Ryan, A. E. 712. The reaction between 3,4,6-tri-O-acetyl-D-glucal and p-nitrophenol. *J. Chem. Soc.* **1962**, 3667.
- (3) Ferrier, R. J.; Zubkov, O. A. Transformation of Glycals into 2,3-Unsaturated Glycosyl Derivatives. *Org. React.* **2003**, *62*, 569.
- (4) Fraser-Reid, B.; Lopez, J. Unsaturated Sugars: A Rich Platform for Methodological and Synthetic Studies. *Curr. Org. Chem.* **2009**, *13* (6), 532–553.
- (5) Bennett, C. S.; Galan, M. C. Methods for 2-Deoxyglycoside Synthesis. *Chem. Rev.* **2018**, *118* (17), 7931–7985.
- (6) Bokor, E.; Kun, S.; Goyard, D.; Toth, M.; Praly, J. P.; Vidal, S.; Somsak, L. C-Glycopyranosyl Arenes and Hetarenes: Synthetic Methods and Bioactivity Focused on Antidiabetic Potential. *Chem. Rev.* **2017**, *117* (3), 1687–1764.
- (7) Ferrier, R. J.; Prasad, N. Unsaturated carbohydrates. Part IX. Synthesis of 2,3-dideoxy- α -D-erythro-hex-2-enopyranosides from tri-O-acetyl-D-glucal. *J. Chem. Soc. C* **1969**, *0* (4), 570–575.
- (8) Bhuma, N.; Lebedel, L.; Yamashita, H.; Shimizu, Y.; Abada, Z.; Ardá, A.; Désiré, J.; Michelet, B.; Martin-Mingot, A.; Abou-Hassan, A.; Takumi, M.; Marrot, J.; Jiménez-Barbero, J.; Nagaki, A.; Blériot, Y.; Thibaudeau, S. Insight into the Ferrier rearrangement by combining flash chemistry and superacids. *Angew. Chem., Int. Ed.* **2020**, DOI: 10.1002/anie.202010175.
- (9) Lebedel, L.; Ardá, A.; Martin, A.; Désiré, J.; Mingot, A.; Aufiero, M.; Aiguabella Font, N.; Gilmour, R.; Jiménez-Barbero, J.; Blériot, Y.; Thibaudeau, S. Structural and Computational Analysis of 2-Halogeno-Glycosyl Cations in the Presence of a Superacid: An Expansive Platform. *Angew. Chem., Int. Ed.* **2019**, *58* (39), 13758–13762.
- (10) Mucha, E.; Marianski, M.; Xu, F.-F.; Thomas, D. A.; Meijer, G.; von Helden, G.; Seeberger, P. H.; Pagel, K. Unravelling the structure of glycosyl cations via cold-ion infrared spectroscopy. *Nat. Commun.* **2018**, *9* (1), 4174.
- (11) Elferink, H.; Severijnen, M. E.; Martens, J.; Mensink, R. A.; Berden, G.; Oomens, J.; Rutjes, F.; Rijs, A. M.; Boltje, T. J. Direct Experimental Characterization of Glycosyl Cations by Infrared Ion Spectroscopy. *J. Am. Chem. Soc.* **2018**, *140* (19), 6034–6038.
- (12) Marianski, M.; Mucha, E.; Greis, K.; Moon, S.; Pardo, A.; Kirschbaum, C.; Thomas, D. A.; Meijer, G.; von Helden, G.; Gilmore, K.; Seeberger, P. H.; Pagel, K. Remote Participation during Glycosylation Reactions of Galactose Building Blocks: Direct Evidence from Cryogenic Vibrational Spectroscopy. *Angew. Chem., Int. Ed.* **2020**, *59* (15), 6166–6171.
- (13) Hansen, T.; Elferink, H.; van Hengst, J. M. A.; Houthuijs, K. J.; Remmerswaal, W. A.; Kromm, A.; Berden, G.; van der Vorm, S.; Rijs, A. M.; Overkleef, H. S.; Filippov, D. V.; Rutjes, F.; van der Marel, G. A.; Martens, J.; Oomens, J.; Codee, J. D. C.; Boltje, T. J. Characterization of glycosyl dioxolenium ions and their role in glycosylation reactions. *Nat. Commun.* **2020**, *11* (1), 2664.
- (14) Greis, K.; Mucha, E.; Lettow, M.; Thomas, D. A.; Kirschbaum, C.; Moon, S.; Pardo-Vargas, A.; von Helden, G.; Meijer, G.; Gilmore, K.; Seeberger, P. H.; Pagel, K. The Impact of Leaving Group Anomerism on the Structure of Glycosyl Cations of Protected Galactosides. *ChemPhysChem* **2020**, *21* (17), 1905–1907.
- (15) Schöllkopf, W.; Gewinner, S.; Junkes, H.; Paarmann, A.; von Helden, G.; Bluem, H. P.; Todd, A. M. M. The new IR and THz FEL facility at the Fritz Haber Institute in Berlin. *Proc. SPIE* **2015**, 95121L.
- (16) Supady, A.; Blum, V.; Baldauf, C. First-Principles Molecular Structure Search with a Genetic Algorithm. *J. Chem. Inf. Model.* **2015**, *55* (11), 2338–48.
- (17) Tkatchenko, A.; Scheffler, M. Accurate molecular van der Waals interactions from ground-state electron density and free-atom reference data. *Phys. Rev. Lett.* **2009**, *102* (7), No. 073005.
- (18) Blum, V.; Gehrke, R.; Hanke, F.; Havu, P.; Havu, V.; Ren, X.; Reuter, K.; Scheffler, M. Ab initio molecular simulations with numeric atom-centered orbitals. *Comput. Phys. Commun.* **2009**, *180* (11), 2175–2196.
- (19) Adamo, C.; Barone, V. Toward reliable density functional methods without adjustable parameters: The PBE0 model. *J. Chem. Phys.* **1999**, *110* (13), 6158–6170.
- (20) Grimme, S.; Antony, J.; Ehrlich, S.; Krieg, H. A consistent and accurate ab initio parametrization of density functional dispersion correction (DFT-D) for the 94 elements H-Pu. *J. Chem. Phys.* **2010**, *132* (15), 154104.
- (21) Frisch, M. J.; et al. *Gaussian 16*, rev. A.03; Gaussian, Inc.: Wallingford, CT, 2016.
- (22) Cremer, D.; Pople, J. A. General definition of ring puckering coordinates. *J. Am. Chem. Soc.* **1975**, *97* (6), 1354–1358.
- (23) Hill, A. D.; Reilly, P. J. Puckering coordinates of monocyclic rings by triangular decomposition. *J. Chem. Inf. Model.* **2007**, *47* (3), 1031–5.